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ORIGINAL PAPER

Cognitive skills and bacterial load: comparative evidence of costs of cognitive proficiency in birds

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Abstract Parasite-mediated selection may affect the evolution of cognitive abilities because parasites may influence development of the brain, but also learning capacity. Here, we tested some predictions of this hypothesis by analyzing the relationship between complex behaviours (feeding innovations (as a measure of behavioural flexibility) and ability to detect foreign eggs in their nests (i.e. a measure of discriminatory ability)) and abundance of microorganisms in different species of birds. A positive relationship would be predicted if these cognitive abilities implied a larger number of visited environments, while if these skills favoured detection and avoidance of risky environments, a negative relationship would be the prediction. Bacterial loads of eggshells, estimated for mesophilic and potentially pathogenic bacteria (i.e. Enterococcus, Staphylococcus and Enterobacteriaceae), were used as a surrogate of probability of contact with pathogenic bacteria. We found that bird species with higher feeding innovation rates and rejection

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M. Martín-Vivaldi Departamento de Biología Animal, Facultad de Ciencias, Universidad de Granada, Granada, Spain rates of experimental brood parasitic eggs had higher density of bacteria on their eggshells than the average species. Since the analysed groups of microorganisms include pathogenic bacteria, these results suggest that both feeding innovation and ability to recognize foreign eggs are costly and highlight the importance of parasite-mediated selection in explaining the evolution of cognitive abilities in animals.

Keywords Behavioural flexibility · Bacterial infection · Brain size · Brood parasitism · Egg rejection · Eggshell bacterial load · Feeding innovation

Introduction

Cognition, which refers to information-processing abilities including perception, learning, memory, judgment and

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91405 Orsay Cedex, France problem solving, has been related to the exploration of novel environments, detection of resources, escape from predators and parasites, and detection of antagonists by prey or hosts implies enormous advantages. These abilities have usually been related to forebrain size and degree of innovation (i.e. technical and feeding innovations), and a relationship between these two estimates has been found in birds (Overington et al. 2009). For example, feeding innovation is related to tool usage and learning in birds and primates (Lefebvre et al. 2002; Nicolakakis and Lefebvre 2000; Lefebvre et al. 2004), and it predicts success of invasive species (Sol et al. 2002, 2005; Sol and Lefebvre 2000). Moreover, a large literature on mammals, including humans, has shown that different kinds of parasitemia (i.e. blood parasite infections) are negatively related to discrimination learning and spatial and non-spatial cognition (review in Garamszegi et al. 2007). Hosts may detect and avoid locations or environments with high risk of parasitism. Moreover, foraging innovation may allow hosts to encounter beneficial substances that help them eliminate parasitism through self-medication (Lozano 1998) or by using material with antibiotic properties for nest building, thereby protecting their offspring (Clark and Mason 1985, 1988; Mennerat et al. 2009). One of the clearest examples of cognitive ability of hosts affecting the interaction with parasites derives from the brood parasitism literature. A main defence of avian hosts against brood parasitism is recognition and rejection of foreign eggs from their nests (Payne 1977; Davies and Brooke 1989; Rothstein 1990). Foreign egg recognition by parasitized birds has evolved as a consequence of selection due to brood parasitism (Soler and Møller 1990; Soler et al. 1999a) and involves cognitive abilities since it implies at least three cognitive tasks (Rothstein 1975b; Lotem et al. 1995; Avilés and Garamszegi 2007): (1) the capacity to discriminate parasitic eggs from own eggs (i.e. judgment), (2) the ability of rejection (i.e. mental decisions related to ejection or burial of parasitic eggs and/or nest desertion), and (3) a linking motivational mechanism that activates rejection once the parasite egg is recognized (i.e. learning of egg appearance). Thus, most evidence suggests important fitness advantages associated with enhanced cognitive proficiency of animals.

Given the hypothetical great advantages associated with increased cognitive ability of animals, one may expect cognitive abilities to increase during evolution. However, this expectation has little support. Variation in cognitive ability within the phylogenetic tree of animals is considerable (Sol et al. 2005), suggesting the existence of phylogenetic costs associated with the evolution of cognition. Improving cognitive ability may increase probability of parasitism if, for instance, it affects the number of different environments that animals exploit. An increased spectrum of visited and explored habitats may also imply encounters with a broader spectrum of disease and infection (Møller and Erritzøe 1998), and thus, risk of parasitism could constrain the evolution of cognitive abilities. On the other hand, increased probability of parasitism could lead to enhanced cognition because individuals with superior cognitive abilities to detect and avoid parasitism would be at a selective advantage. Both scenarios predict a relationship between parasitism and cognitive ability of animals (Garamszegi et al. 2007; Fig. 1). Here, we suggest that bacteria may play an important role in this scenario and limit the evolution of cognition.

Bacteria are ubiquitous microorganisms that, when colonizing animal tissues, may have positive, negative, or neutral effects on fitness (McArthur 2006). In fact, a major part of the immune system (i.e. humoral immunity) has evolved for fighting bacterial infection in general (innate immunity mediated by natural antibodies) and the most commonly encountered pathogenic microorganisms in particular (Playfair and Bancroft 2004; Matson et al. 2005). Environmental conditions determine the distribution of pathogenic bacteria, which are acquired by animal hosts when visiting different environments through direct contact with other individuals, by aquatic or aerial dispersal, through ingestion of food or from biological vectors (Atlas and Bartha 2001). Thus, animals with a higher degree of feeding innovation (i.e. enhanced cognitive abilities) may visit a larger variety of environments and consequently experience a higher risk of pathogenic bacterial infection than others. Moreover, feeding innovation is correlated with other opportunistic behaviours (Nicolakakis and Lefebvre 2000), which may increase the risk of exposure to a wide range of parasites, and therefore, parasite loads should be larger in innovative species (Lefebvre et al. 2002; Fig. 1).



Fig. 1 Hypothetical relationships between cognitive skills (i.e. feeding innovation and rejection of foreign egg), brain size and probability of parasitism (bacterial infection) suggesting a role of parasite-mediate natural selection in the evolution of cognitive proficiency of animals. See text for further details

Feeding innovation reliably reflects an important component of cognitive ability of animals (Sol et al. 2005). In accordance with the hypothesis that risk of parasitism could constrain the evolution of cognitive abilities of animals, Garamszegi et al. (2007) have shown that bird species with higher level of feeding innovation have larger immune defence organs and higher prevalence of Haematozoa. Moreover, Vas et al. (2011) have recently shown that more innovative species are parasitized by a more diverse lice fauna (Amblycera). These two results suggest that the evolution of feeding innovation has implications for parasite-mediated natural selection.

Here, we tested this hypothesis in a comparative framework using both feeding innovation and ability to detect foreign eggs in their nests as proxies of cognitive abilities of birds. Eggshell bacterial loads were used as a proxy of probability of contact with pathogenic bacteria that should affect the risk of infection of embryos that are particularly susceptible to parasitism. Feeding innovation may represent cognitive abilities related to behavioural flexibility, technical innovation and/or ability to exploit novel environments, while rejection rates of parasitic egg indicate the ability of birds to locate deviant objects within their particular nest environment (Rothstein 1975a, 1975b; Lyon 2003; Shizuka and Lyon 2010). The two estimates of cognitive abilities reflect important aspects of cognition in birds. The bacterial environment of bird nests can be estimated by quantifying eggshell bacterial loads. Eggshell bacterial density is positively related to probability of embryo infection (Bruce and Drysdale 1994; Cook et al. 2003, 2005b) that is more likely to occur before or at the beginning of incubation (Cook et al. 2003), and thus, our estimations of eggshell bacterial loads are likely to reflect the probability of embryo infection. Since cognitive skills are related to the activity and the diversity of habitats encountered by adult birds, we predicted a positive relationship between cognitive skills and eggshell bacterial loads. Cognitive abilities are known to be associated with brain size (Sol et al. 2005; Avilés and Garamszegi 2007), and consequently, we included brain size as an additional independent variable in the models. Hypothetical background supporting the predicted relationships between cognitive skills of birds and probability of bacterial infections is shown in Fig. 1. Support for the hypothesis would suggest unexplored costs associated with cognitive skills and would thus have important consequences for understanding the evolution of cognition in animals.

Materials and methods

Study sites and nest locations

Bacterial communities on eggshells of nests were sampled during the breeding seasons 2007–2008 and 2006–2008 at

Kraghede (Denmark, 57° 12' N, 10° 00' E) and Guadix (Spain, $37^{\circ}18$ 'N, $3^{\circ}11$ 'W), respectively. For a detailed description of the Danish and Spanish study area, see, respectively, Møller (1987) and Martín-Vivaldi et al. (2006) and Soler and Avilés (2010).

JMP, EFJ and APM in Denmark and JMP, JJS and MMV in Spain made extensive systematic searches for nests in suitable habitats throughout the breeding season. We relied on extensive help from amateurs with a good knowledge of birds in locating nests of all common breeding species. Adult birds carrying nest material in their beak were particularly used as a means of locating nests with fresh eggs. Most nests were therefore located during nest building by intensively searching suitable habitat in the study area and by checking nest boxes. We deliberately attempted not to touch nests or disturb the surrounding vegetation to avoid increasing the risk of nest predation. When a nest was detected during egg laying, on the basis of the typical clutch size of the species, we estimated the date of clutch completion and visited the nests the following day to sample eggshell bacteria. Nests were re-visited at hatching. The number of nest checks was, in this way, minimized to reduce any unnecessary predation due to investigators.

Estimation of eggshell bacterial loads

We sampled eggs at the beginning of incubation (i.e. 2– 3 days after clutch completion), which assured that independent of the species, all sampled eggs were incubated, and therefore, possible interspecific differences in eggshell bacterial loads due to variation in the onset of incubation (Cook et al. 2003) were partially mitigated.

While sampling eggshells and mainly to prevent between nest contamination, we wore latex gloves sterilized with ethanol and took bacterial samples by cleaning eggshells with a sterile swab slightly wet with sterile sodium phosphate buffer (0.2 M; pH 7.2). The entire clutch was cleaned with the same swab, which was preserved in an eppendorf tube at 4°C containing the sterile buffer until lab analyses during the following 30 days. The duration of the storage period did not affect rank position of different species as shown by a comparison of ranked values of heterotrophic bacterial loads of 21 species from which we, in 2006, collected samples that were stored less than 3 days (N=120) and resulting values of adding other samples that were stored up to one month (N=156) (Kendall Coefficient of Concordance=0.95; average Spearman Rank Correlation= 0.91, Friedmann ANOVA, χ^2 =38.13, P=0.009). Estimates of bacterial load were standardized for total eggshell surface sampled by taking into account the number and the surface of eggs in the nests. Eggshell surface was estimated according to the formula:

$$S = (3.155 - 0.0136 \times L + 0.0115 \times W) \times L \times W$$

(Narushin 2005) where S is the surface in square centimetres, L is the length of the egg and W is the width of the egg. Length and width of all eggs were measured with a calliper (accuracy: 0.02 mm).

In the lab, samples were collected from eppendorf tubes after vigorously shaking the eppendorf in vortex for at least three periods of 5 s. Serial decimal dilutions up to 10^{-6} were cultivated by spreading homogeneously 100 µl of sample (measured with a micropipette) in plates containing four different sterile solid growth media (Scharlau Chemie S.A. Barcelona). We used tryptic soy agar (TSA), a broadly used general medium to grow mesophilic bacteria, and three specific media: Kenner Fecal Agar (KF) for growing bacteria belonging to the genus Enterococcus, Vogel-Johnsson Agar (VJ) for bacteria of the genus Staphylococcus, and Hecktoen Enteric Agar (HK) for Gram-negative bacteria of the family Enterobacteriaceae. Plates were incubated at 32°C for 72 h, and afterwards, the number of colonies on each plate was counted. Bacterial density was estimated as colony forming units (CFU) per square centimetre. See Peralta-Sánchez et al. (2010) for repeatability estimates of intraspecific variation. Permission for egg manipulations were granted from the Junta de Andalucía (Spanish Regional Government).

Enterobacteriaceae and Staphylococcus sp. include saprophytic and opportunistic bacteria (Singleton and Harper 1998; Houston et al. 1997; Cook et al. 2005a) that live on skin, hair and feathers of mammals and birds (Krieg and Holt 1984). They commonly appear on avian eggshells and are known to include pathogenic strains for avian embryos (Bruce and Drysdale 1994). Enterococci, the third analysed group of bacteria, are also frequently found inside unhatched eggs (Bruce and Drysdale 1994) and are opportunistic pathogens (Franz et al. 1999), although some species might also have beneficial effects (Moreno et al. 2003; Soler et al. 2008, 2010; Martín-Vivaldi et al. 2010). Most of these bacteria are able to penetrate eggshells, and therefore, their density predicts probability of embryonic infections (Board et al. 1994; Cook et al. 2003). In addition, eggshell density of bacteria able to grow in aerobic mesophilic medium is positively related to the probability of embryo infection (Bruce and Drysdale 1994; Cook et al. 2003, 2005b). Therefore, there are good reasons for considering the estimated eggshell bacterial loads as proxies for probability of transshell bacterial infection of embryos.

Feeding innovation and rejection rates

Feeding innovations can be quantified from the ornithological literature, using descriptions of novel kinds of feeding behaviour (Lefebvre et al. 1997). This data set was collated from an exhaustive survey of 30 years (1970–2000) of the short note sections of 65 generalist ornithology journals covering six geographical areas of the world (Lefebvre et al.

2004). For a detailed description of the systematic data collection, see Lefebvre et al. (1997, 2001), Nicolakakis and Lefebvre (2000), Sol et al. (2002) and Nicolakakis et al. (2003). These estimates are species-specific attributes, as shown by comparisons of estimates for the same species in different continents (Garamszegi et al. 2007).

For rejection rates of potential hosts of the European cuckoo (Cuculus canorus), we used the weighted mean values calculated for the different experiments with nonmimetic models performed by different scientists throughout Europe (von Haartman 1981; Järvinen 1984; Davies and Brooke 1989; Moksnes et al. 1991; Moksnes et al. 1994; Palomino 1997; Brooke et al. 1998; Alvarez 1999; Moskat and Fuisz 1999; Stokke et al. 1999; Lindholm and Thomas 2000; Grim and Honza 2001; Amundsen et al. 2002; Rutila et al. 2002, 2006; Moskat et al. 2003a; Prochazka and Honza 2003, 2004; Honza et al. 2004; Lovaszi and Moskat 2004; Antonov et al. 2006b; 2006a; 2009; 2010; Stokke et al. 2008), including our data from Sierra Nevada, Spain (Martín-Vivaldi et al. 2012). We only considered studies providing data on rejection rates of artificially introduced non-mimetic models or eggs that did not use additional stimuli reflecting risk of parasitism (cuckoo dummies). Experiments using conspecific eggs with their natural coloration or models or eggs painted to be similar to the specific gens (host race) of the host species being tested are not included. Rejection rates of non-mimetic model eggs have been broadly used in the literature as a surrogate of recognition ability of foreign eggs by potential hosts of brood parasites (Avilés and Garamszegi 2007; Soler and Møller 1996; Soler et al. 1999b; Soler 1999). However, the single study that has attempted to relate egg-rejection rates to variables reflecting cognitive abilities (i.e. relative brain size) found that species with relatively larger brains had lower egg-rejection rates (Avilés and Garamszegi 2007). However, total brain size may not appropriately reflect the ability of birds to discriminate against foreign eggs in their nests because particular tasks may be more closely related to particular areas of the brain (Timmermans et al. 2000). In any case, we statistically controlled the predicted relationship between egg-rejection rates and bacterial loads of eggshells by inclusion of relative brain size of birds as an additional predictor variable (see the following discussion).

Information on feeding innovation and rejection rate is shown in Online Resource 1.

Brain size

Information on brain size was obtained from data on brain mass reported by Mlikovsky (1989), Iwaniuk and Nelson (2003) and J. Erritzøe (personal communication). Highly significant repeatabilities among studies indicate that information on brain mass can be combined across sources (Garamszegi et al. 2005). Relative brain mass was obtained from the residuals of the log–log regression of brain mass against body mass. Brain mass and body mass values are given in Online Resource 1. This relationship between brain mass and body mass had a strong phylogenetic signal (λ = 0.93), and thus, residuals were estimated from the phylogenetic generalized least square regression model (Pagel 1997, 1999) as implemented in R with the appropriate libraries ("ape", "MASS" and "mvtnorm") and additional unpublished functions by R. Freckleton (University of Sheffield) (pglm3.3.r, available on request) as implemented in the package "caic" and using the composite molecular phylogeny described in the following discussion and shown in Online Resource 1.

Sample sizes and statistical analyses

We successfully collected information on eggshell bacterial loads for 1,012 nests (346 from Denmark and 666 from Spain) of 58 species of birds (49 from Denmark and 25 from Spain) (see Online Resource 1). For 55 of these species, information on feeding innovation was available, while information on rejection rates of non-mimetic model eggs was available for 25 species. For 22 species with estimates of eggshell bacterial loads, we obtained information on both feeding innovation and rejection rates (see Online Resource 1).

We know from previous analyses that interspecific variation in eggshell bacterial load is significantly larger than intraspecific and intervear variation (Peralta-Sánchez 2011) (results from GLM's testing for effects of year (random factor) and species identity (fixed factor) on rank-transformed eggshell bacterial density estimated at the beginning of incubation in four different growth media; $F_{18, 538} > 6.5$, P < 0.001), and thus, we estimated geometric means of log₁₀-transformed eggshell bacterial load for each species sampled in each country. Frequency distributions of log10-bacterial density did not differ from normality for mesophilic bacteria or Enterobacteria (Kolmogorov-Smirnov tests for continuous variables, P> 0.2), but did so for Enterococci and Staphylococcus (Kolmogorov–Smirnov tests for continuous variables, P < 0.05). Thus, because we were interested in considering all estimations of eggshell bacterial load in the same analyses, we ranked values of eggshell bacterial loads and used these in subsequent analyses. Moreover, because we were not interested in differences among locations, but in maximizing the number of species with information on eggshell bacterial loads, we used residuals of such values after controlling for the effect of country. For species sampled in both countries, we used mean values of these residuals. Residuals from the statistical models did not differ from normal distributions (Kolmogorov-Smirnov test for continuous variables, p > 0.2), which justify the use of parametric statistical tests.

The probability of finding a feeding innovation in different species may depend on several methodological factors, and therefore, absolute counts of innovation events should be corrected (Lefebvre et al. 1997; Sol et al. 2002). First, there may be more reports available for intensely studied species. We estimated research effort by using the number of studies published since 1972 on each species as cited in the ISI Web of Science (http://www.isiknowledge.com/). Second, we assessed the importance of population size that may affect the probability of detection of feeding innovations. We used the minimum estimates of breeding population size (in number of pairs) given in Tucker and Heath (1994), rounding population size to the nearest million if more than a million pairs were found for a species. Third, we assessed the potential confounding effect of body mass on feeding innovations because larger species may be more likely observed when feeding. We used our own data for body mass (see Garamszegi et al. 2007). Trying to correct our analyses for these potential confounding factors, we included them in the MANOVA models explaining eggshell bacterial loads and removed those with the largest (non-significant) P-values one by one. After removing population size (Wilks=0.90, $F_{4.46}$ =1.34, P=0.27) and body mass (Wilks=0.87, F_{4,47}=1.70, P=0.17), final models included estimates of research effort and feeding innovation. Thus, we present results from these final models. The most common and abundant species may be those with higher research effort and more frequent feeding innovation, and thus, we repeated the analyses without including research effort in the models. Brain size relative to body size of birds is known to be positively (Sol et al. 2005) and negatively (Avilés and Garamszegi 2007) related to rates of feeding innovation and foreign egg rejection, respectively, and thus, models including residual brain size after controlling for body mass were also run.

Interspecific statistical relationships could be affected by phylogenetic non-independence (Harvey and Pagel 1991). Therefore, we tested residuals of the regressions between estimated eggshell bacterial loads and feeding innovation and rejection rates using the lambda statistic of Pagel (1999; Freckleton et al. 2002), relying on a composite molecular phylogeny derived from Jønsson and Fjeldså (2006), Hackett et al. (2008) and Pons et al. (2005) with branch length arbitrarily assigned to one (Online Resource 1). We found that the estimated values of λ were either not statistically significantly different from zero (relationships between rejection rates and eggshell density of Staphylococcus and between eggshell density of Enterobacteriaceae and feeding innovation and research effort) or equal to zero (all other analysed relationships) (see Online Resource 1), indicating that there was no phylogenetic signal, and therefore, the analyses did not need to be corrected for phylogenetic effects (see, Sæther et al. 2011).

The predicted relationships between eggshell bacterial loads and avian cognitive abilities were tested by means of MANOVAs with eggshell bacterial loads (i.e. mesophilic bacteria, *Enterococcus*, *Staphylococcus* and *Enterobacteriaceae*) as dependent variables and rejection rate and feeding innovation rate (and research effort for correcting feeding innovation) as independent continuous variables in the models. These MANOVAs were weighted by the number of nests of each species that were sampled for eggshell bacterial load estimations which should adjust our comparative analyses for heterogeneity in data quality due to the large variation in sample sizes among species (Garamszegi and Møller 2010).

Results

Estimates of rejection and feeding innovation rates were not significantly related (*Beta* (*SE*)=0.04 (0.26), $F_{1,20}$ =0.04, P=0.85), and thus, they represent two different measures of cognitive abilities.

Potential host species of the European cuckoo showed higher rejection rates of non-mimetic model eggs if they had higher bacterial loads on their eggshells (weighted MAN-OVA, Wilks=0.355, F_{4,20}=9.09, P=0.0002). Univariate results indicate that this association was mainly due to the positive association with mesophilic bacteria (Beta (SE)= 0.63 (0.16), P=0.0007), Enterococci (Beta (SE)=0.43 (0.19), P=0.030) and Enterobacteria (Beta (SE)=0.47) (0.18), P=0.017) (Fig. 2), while no significant association was detected for Staphylococci (Beta (SE)=-0.21 (0.20), P= 0.31). Inclusion of residual brain mass in the model after correction for body mass and phylogeny did not affect the main conclusions (weighted MANOVA, effect of rejection rates: Wilks=0.413, F_{4,19}=6.74, P=0.0014; effect of brain mass: Wilks=0.700, F_{4,19}=2.08, P=0.124). Univariate results of this model confirmed the relationship between rejection rates and abundance of mesophilic bacteria (Beta (SE)=0.46 (0.15), P=0.006), but not that with Enterococci (Beta (SE)=0.40 (0.21), P=0.062) and Enterobacteriaceae (Beta (SE)=0.30 (0.18), P=0.110), while brain mass was positively associated with abundance of mesophilic bacteria (Beta (SE)=0.45 (0.15), P=0.001) and Enterobacteriaceae (Beta (SE)=0.45 (0.18), P=0.022). No other bacterial counts were significantly related with rejection rates or brain mass (P>0.18).

Species with higher rates of feeding innovation had larger eggshell bacterial loads (weighted MANOVA, Wilks= 0.617, $F_{4,50}$ =7.76, P=0.0009). Univariate results showed that this relationship was due to the positive association between eggshell density of *Staphylococcus* and feeding innovation (*Beta* (*SE*)=0.51 (0.19), P=0.0001; Fig. 3) because no other bacterial count showed a significant effect (P>0.45). Inclusion in the model of the statistically significant effects of brain mass (weighted MANOVA, Wilks=0.685, $F_{4,48}$ =5.51,

P=0.001) did not affect the strength of the detected effect of feeding innovation on eggshell bacterial loads (weighted MANOVA, Wilks=0.704, $F_{4,48}=5.06$, P=0.0018). Univariate results of this model showed that eggshell density of



Fig. 2 Relationship between eggshell bacterial loads of potential host species of the European cuckoo estimated for mesophilic bacteria (\mathbf{a}), Enterococci (\mathbf{b}) and Enterobacteriaceae (\mathbf{c}), and rejection rates of non-mimetic model eggs. *Lines* are weighted regression lines. The areas of *circles* are proportional to log-transformed number of nests of each species sampled for eggshell bacterial loads



Fig. 3 Relationship between eggshell bacterial loads of potential host species of the European cuckoo estimated for Staphylococci bacteria and feeding innovations. *Lines* are weighted regression lines. The areas of *circles* are proportional to log-transformed number of nests of each species sampled for eggshell bacterial loads

Staphylococcus was positively related to feeding innovation (Beta (SE)=0.65 (0.14), P=0.0001) and negatively related to brain mass (Beta (SE)=-0.39 (0.14), P=0.01). In addition, residual brain mass was negatively related to Enterobacteriaceae on eggshells (Beta (SE)=-0.47 (0.15), P=0.003), while no other univariate results reached statistical significance (P>0.06). When we included research effort (log-transformed) for estimating feeding innovation in the previous model, neither research effort (weighted MANOVA, Wilks=0.856, $F_{4,47}$ = 1.98, P=0.113) nor feeding innovation did explain a significant proportion of interspecific variance in eggshell bacterial loads (weighted MANOVA, Wilks=0.851, $F_{4,47}$ =2.05, P= 0.102), but residual brain mass did (weighted MANOVA, Wilks=0.763, *F*_{4,47}=3.65, *P*=0.011). However, univariate results confirmed that feeding innovation tended to explain density of Staphylococci (Beta (SE)=0.44 (0.23), P=0.056), while brain size was negatively related to density of both

Table 1 Results from MANOVA testing the relationship between bacterial loads of eggshells estimated as density of mesophilic bacteria (TSA), Enterococci (KF), Staphylococci (VJ) and Enterobacteria (HK) as dependent factors, and feeding innovations (log-transformed), research effort (log-transformed) for estimates of feeding innovation,

Staphylococci (*Beta* (*SE*)=-0.33 (0.15), P=0.037) and *Enter*obacteriaceae (*Beta* (*SE*)=-0.45 (0.17), P=0.009). The relationships between research effort for estimating feeding innovation and eggshell bacterial loads were far from significant independently of the specific bacterial count (P>0.22).

Finally, when including information on rejection rates and feeding innovation in the same model, thereby greatly reducing sample size, the conclusions remained unchanged. Both feeding innovation (weighted MANOVA, Wilks=0.374, $F_{4.16}$ =6.70, P=0.002) and rejection rates (weighted MAN-OVA, Wilks=0.287, F_{4.16}=9.93, P=0.0003) were significantly positively associated with eggshell bacterial load. Univariate results indicated that two estimates of eggshell bacterial density were significantly related to rejection rates (mesophilic bacteria: Beta (SE)=0.62 (0.18), P=0.003; Enterobacteria (Beta (SE)=0.46 (0.20), P=0.035)), while that was not the case for Enterococci (Beta (SE)=0.41 (0.20), P=0.052) and Staphylococci (Beta (SE)=-0.21 (0.15), P=0.18). The relationship with feeding innovation was mainly due to the positive association with Staphylococcus load (Beta (SE)=0.73 (0.15), P=0.0001, with no other bacterial count being significantly related to feeding innovation rate (results not shown).

When residual brain mass and sampling effort for feeding innovation estimates were included as additional factors in the model, rejection rate but not feeding innovation was still significantly associated with eggshell bacterial load, and both research effort and residual brain mass entered significantly in the model (Table 1). Thus, it is possible that lack of statistical significance for feeding innovation was due to research effort and feeding innovation sharing a considerable amount of variance (Garamszegi et al. 2007). Univariate results confirmed the positive associations between variables related to cognitive skills and eggshell bacterial loads since the abundance of Staphylococci was positively related to feeding innovation and research effort, while rejection rate was related to Enterococci. Residual brain mass were positively related to density of mesophilic bacteria (Table 1).

residual brain size after controlling for body mass (Brain size) and rejection rates of non-mimetic model eggs (square-root arcsinetransformed) as predictors. Multivariate (Wilks and associated *F*-statistics) and univariate results (Beta, standard error (SE) and associated *P*-value) for each kind of quantified bacteria are shown

				e	,	,		1			
				Univariate results							
				TSA		KF		VJ		НК	
	Wilks	$F_{4,14}$	Р	Beta (SE)	Р	Beta (SE)	Р	Beta (SE)	Р	Beta (SE)	Р
Feeding innovation	0.623	2.121	0.132	0.06(0.21)	0.78	0.02(0.26)	0.95	0.43(0.19)	0.039	-0.03(0.25)	0.89
Research effort	0.387	5.533	0.007	-0.21(0.24)	0.40	0.46(0.30)	0.14	0.48(0.22)	0.040	0.08(0.28)	0.77
Brain size	0.427	4.689	0.013	0.53(0.18)	0.009	-0.12 (0.23)	0.60	0.01(0.17)	0.96	0.45(0.22)	0.053
Rejection rate	0.492	3.610	0.033	0.62(0.21)	0.13	0.65(0.26)	0.023	0.01(0.19)	0.97	0.33(0.25)	0.20

When relative brain size was the only independent variable included in the model, it explained a significant proportion of interspecific variance in eggshell bacterial load (weighted MANOVA, Wilks=0.619, $F_{4,63}$ =9.69, P=0.00004). This association was mainly due to the significant negative associations with Enterococci (*Beta* (*SE*)=-0.38 (0.11), *P*=0.001) and *Enterobacteriaceae* (*Beta* (*SE*)=-0.45 (0.11), *P*= 0.0001), while no other bacterial count was significantly associated with brain size (*P*>0.06). These results indicate that, in general, birds with larger brain size enjoy reduced probability of eggshell bacterial infection.

Discussion

Previously, Garamszegi et al. (2007) and Vas et al. (2011) found a positive interspecific relationship between feeding innovation and prevalence of haematozoan parasitism and diversity of parasitic Amblyceran lice, respectively. These results could be interpreted in a general scenario if feeding innovation reflected complex cognitive functions. Behavioural flexibility and opportunism may be associated with environmental variability, width of the ecological niche, social structure, morphological variability and population density (Lefebvre and Bolhuis 2003; Vas et al. 2011), which may all involve increased parasite-mediated selection. However, feeding innovation per se may increase the risk of parasitism because, independently of other cognitive skills, opportunistic feeding behaviour by exploiting novel food sources and/or acquiring novel feeding styles may imply higher probability of encountering pathogenic microorganisms. Therefore, Garamszegi et al. (2007) concluded that "further analyses are required to distinguish whether feeding innovation alone or its cognitive or ecological correlates enhance the risk of parasitism". Here, we used two different independent estimates of cognitive proficiency of animals, feeding innovation and ability to recognize foreign eggs and found an association with intensity of bacterial parasitism, inferred from estimates of eggshell bacterial loads. Thus, our results lend further support to the existence of evolutionary costs of being innovative.

Cognitive abilities and costs of microbial infection

Costs associated with cognitive abilities of animals are not yet well understood. Most hypothetical costs are mainly energetic (Aiello and Wheeler 1995; Sengupta et al. 2010). In accordance with the idea that cognitive abilities may trade against energy consumption (Aiello and Wheeler 1995), there is evidence suggesting that larger brain size is traded against the size of other energetically expensive tissues (reviewed in Niven and Laughlin 2008). Interestingly, resolution of this trade-off would vary depending on environmental conditions (i.e. habitat) and life history traits, which therefore may explain extreme variation in cognitive proficiency among animals (Burns et al. 2011). The probability of parasitism also varies depending on environmental conditions (Møller et al. 2006; Merino and Potti 1996), and it strongly affects the evolution of life history traits of animals (Møller 1997; Martin et al. 2001). Consequently, parasites could also play a role in the resolution of trade-offs between cognitive skills and energy consuming activities. For example, it is known that honey bees Apis melifera (Mallon et al. 2003) and bumblebees Bombus terrestris (Riddell and Mallon 2006) perform poorly in learning assays when their immune system has been challenged by a non-pathogenic elicitors of the immune system. Apart from a trade-off between energy consuming activities (Alghamdi et al. 2009), this association may also suggest a role of parasites driving the evolution of cognitive abilities if animals with enhanced cognitive proficiency experienced higher probability of parasitism, as is likely to be the case (i.e. larger number of flowers visited; Ruiz-Gonzalez and Brown 2006). In this situation, individuals or species with high probability of parasitism (higher cognitive abilities) should be those with stronger anti-parasite defences (Møller and Erritzøe 2002). Therefore, this scenario predicts a positive association between immune response and cognitive proficiency in birds (Garamszegi et al. 2007) and bumblebees (Alghamdi et al. 2009). We have found a positive association between two different components of cognitive abilities and probability of bacterial infection, which further suggests an important role of parasitism in driving the evolution of cognition in animals.

We used brain size in our analyses to statistically control the association between cognitive tasks (feeding innovation and foreign egg rejection) and bacterial infection. After correcting for rejection rates, species with larger brains had higher bacterial density. However, after correcting for rate of feeding innovations, species with larger brain size had lower bacterial density on their eggshells. Estimates of rejection and feeding innovation rates were not significantly related to each other, and the association between brain size and rejection rate was negative, while the association between brain size and feeding innovation rate was positive. Finally, relative brain size was negatively related to eggshell bacterial load when no other independent variables were included in the statistical models. Variation in the sign of the association between brain mass and eggshell bacterial load depending on the independent variables included in the models should obviously be interpreted as partial correlation coefficients. Consequently, these results suggest, first, that birds with relatively larger brains have higher capacity of eluding bacterial contamination of eggshells and, second, that for species with similar relative brain size, those with higher rates of feeding innovation and egg rejection had higher bacterial density on eggshells. Relative brain size has repeatedly been used as indicative of cognitive capacity of animals (Lefebvre et al.

1997, 2004; Sol et al. 2005), and thus, these results could be interpreted as brain size playing a role in diminishing bacterial infection, while bird species with higher cognitive activities relative to their brain size suffer higher risk of bacterial infection.

Recognition of brood parasitic eggs and costs of microbial infection

Particularly intriguing is the association between rejection ability of potential hosts of the common cuckoo and eggshell bacterial density since it would represent unexplored costs associated with recognition of foreign eggs that would help to explain the low or intermediate rates of egg rejection of most suitable hosts (Rothstein 1990). The study of costs of egg rejection has focused on the existence of recognition errors (Lotem et al. 1995; Davies et al. 1996), costs of misimprinting on parasitic eggs (Lotem 1993) or breakage of own eggs when trying to eject parasitic eggs (Rohwer et al. 1989; Lotem et al. 1995; Martín-Vivaldi et al. 2002). Our analyses show an increased probability of bacterial infections associated with superior egg-recognition skills, suggesting an important additional cost constraining the evolution of rejection ability. This relationship may suggest that cognitive skills of hosts of brood parasites when encountering a parasitic egg in their nests may be non-adaptive in other environments in which it increases the probability of bacterial infection. Egg recognition proficiency of birds may indicate neophilia and ability to find strange objects in particular environment (Rothstein 1975a, 1975b; Lyon 2003; Shizuka and Lyon 2010). It is believed that the ability to recognize foreign eggs evolved as a consequence of favourable cognitive skills related to nest sanitation (Bartol et al. 2003; Moskat et al. 2003b; Guigueno and Sealy 2009). Thus, it is unlikely that the positive relationship with eggshell bacterial loads were related to a lower effort maintaining hygienic conditions of nests by bird species with higher rejection rates. Rather, these cognitive skills may make birds more prone to explore environments, objects or locations with an associated increased probability of encountering bacteria. However, we did not find support for a positive relationship between foreign-egg recognition ability and feeding innovation. These suggestions should, in any case, be experimentally tested to reach further conclusions. An alternative explanation for the detected relationship between egg-rejection ability and eggshell bacterial load of potential host species is related to the historical associations with the European cuckoo. For instance, adults of hosts and parasitic cuckoos differ in their intestinal microbiota (see Ruiz-Rodriguez et al. 2009), and consequently, the addition of brood parasitic eggs to the host clutch may have resulted in a more diverse bacterial community of the eggs of the most commonly parasitized hosts. Moreover, some rejecter species puncture parasitic eggs or break some of their own eggs (Soler et al.

2002; Antonov et al. 2006c), which may deteriorate the hygienic conditions in host nests (Soler et al. 2011). Nest lining material and eggs could become smudged with yolk and egg white from damaged eggs, increasing nutrient availability for bacterial growth on the eggshell (Stadelman 1994). Although we cannot reject these alternative scenarios explaining the detected interspecific association between egg-rejection ability and eggshell bacterial loads, none of the sampled nests were parasitized by the European cuckoo, and thus, these alternatives are improbable.

In summary, we found a positive relationship between intensity of bacterial colonization of avian eggshells and two variables reflecting different cognitive abilities of birds, rates of feeding innovation and recognition of foreign eggs. These relationships suggest that parasite-mediated selection may constrain the evolution of cognitive proficiency in birds. Studies at the intraspecific level are needed to further examine the causes explaining the relationships detected here.

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References

- Aiello LC, Wheeler P (1995) The expensive-tissue hypothesis: the brain and the digestive system in human and primate evolution. Curr Anthropol 36:199–221
- Alghamdi A, Raine NE, Rosato E, Mallon EB (2009) No evidence for an evolutionary trade-off between learning and immunity in a social insect. Biol Lett 5:55–57
- Alvarez F (1999) Attractive non-mimetic stimuli in cuckoo *Cuculus* canorus eggs. Ibis 141:142–144
- Amundsen T, Brobakken PT, Moksnes A, Røskaft E (2002) Rejection of common cuckoo Cuculus canorus eggs in relation to female age in the bluethroat Luscinia svecica. J Avian Biol 33:366–370
- Antonov A, Stokke BG, Moksnes A, Røskaft E (2006a) Coevolutionary interactions between common cuckoos and corn buntings. Condor 108:414–422
- Antonov A, Stokke BG, Moksnes A, Røskaft E (2006b) Egg rejection in marsh warblers (*Acrocephalus palustris*) heavily parasitized by common cuckoos (*Cuculus canorus*). Auk 123:419–430
- Antonov A, Stokke BG, Moksnes A, Kleven O, Honza M, Røskaft E (2006c) Eggshell strength of an obligate brood parasite: a test of the puncture resistance hypothesis. Behav Ecol Sociobiol 60:11– 18
- Antonov A, Stokke BG, Moksnes A, Røskaft E (2009) Evidence for egg discrimination preceding failed rejection attempts in a small cuckoo host. Biol Lett 5:169–171
- Antonov A, Stokke BG, Ranke PS, Fossøy F, Moksnes A, Røskaft E (2010) Absence of egg discrimination in a suitable cuckoo Cuculus canorus host breeding away from trees. J Avian Biol 41:501– 504
- Atlas RM, Bartha R (2001) Ecología microbiana y microbiología ambiental. Addison Wesley Longman, Inc., Madrid

- Avilés JM, Garamszegi LZ (2007) Egg rejection and brain size among potential hosts of the common cuckoo. Ethology 113:562–572
- Bartol I, Moskat C, Karcza Z, Kisbenedek T (2003) Great reed warblers bury artificial objects, not only cuckoo eggs. Acta Zool Acad Sci Hung 49:111–114
- Board RG, Clay C, Lock J, Dolman J (1994) The egg: a compartmentalized, aseptically packaged food. In: Board RG, Fuller R (eds) Microbiology of the avian egg. Chapman & Hall, London, pp 43– 62
- Brooke MdL, Davies NB, Noble DG (1998) Rapid decline of host defenses in response to reduced cuckoo parasitism: behavioral flexibility of reed warblers in a changing world. Proc R Soc Lond B 265:1277–1282
- Bruce J, Drysdale EM (1994) Trans-shell transmission. In: Board RG, Fuller R (eds) Microbiology of avian eggs. Chapman & Hall, London, pp 63–91
- Burns JG, Foucaud J, Mery F (2011) Costs of memory: lessons from "mini' brains. Proc R Soc B. doi:10.1098/rspb.2010.2488:
- Clark L, Mason JR (1985) Use of nest material as insecticidal and antipathogenic agents by the European starling. Oecologia 67:169–176
- Clark L, Mason JR (1988) Effect of biologically active plants used as nest material and the derived benefit to starling nestlings. Oecologia 77:174–180
- Cook MI, Beissinger SR, Toranzos GA, Rodriguez RA, Arendt WJ (2003) Trans-shell infection by pathogenic micro-organisms reduces the shelf life of non-incubated bird's eggs: a constraint on the onset of incubation? Proc R Soc Lond B 270:2233–2240
- Cook MI, Beissinger SR, Toranzos GA, Arendt WJ (2005a) Incubation reduces microbial growth on eggshells and the opportunity for trans-shell infection. Ecol Lett 8:532–537
- Cook MI, Beissinger SR, Toranzos GA, Rodriguez RA, Arendt WJ (2005b) Microbial infection affects egg viability and incubation behavior in a tropical passerine. Behav Ecol 16:30–36
- Davies NB, Brooke MdL (1989) An experimental study of coevolution between the cuckoo, *Cuculus canorus*, and its host. I. Host egg discrimination. J Anim Ecol 58:207–224
- Davies NB, MdL B, Kacelnik A (1996) Recognition errors and probability of parasitism determine whether reed warblers should accept or reject mimetic cuckoo eggs. Proc R Soc Lond B 263:925–931
- Franz CMAP, Holzapfel WH, Stiles ME (1999) Enterococci at the crossroads of food safety? Int J Food Microbiol 47:1–24
- Freckleton RP, Harvey PH, Pagel M (2002) Phylogenetic analysis and comparative data: a test and review of evidence. Am Nat 160:712–726
- Garamszegi LZ, Møller AP (2010) Effects of sample size and intraspecific variation in phylogenetic comparative studies: a metaanalytic review. Biol Rev 4:797–805
- Garamszegi LZ, Erritzøe J, Møller AP (2007) Feeding innovations and parasitism in birds. Biol J Linn Soc 90:441–455
- Garamszegi LZ, Eens M, Erritzoe J, Møller AP (2005) Sexually size dimorphic brains and song complexity in passerine birds. Behav Ecol 16:335–345
- Grim T, Honza M (2001) Differences in behaviour of closely related thrushes (*Turdus philomelos* and *T merula*) to experimental parasitism by the common cuckoo *Cuculus canorus*. Biologia 56:549–556
- Guigueno MF, Sealy SG (2009) Nest sanitation plays a role in egg burial by yellow warblers. Ethology 115:247–256
- Hackett SJ, Kimball RT, Reddy S, Bowie RCK, Braun EL, Braun MJ, Chojnowski JL, Cox WA, Han KL, Harshman J, Huddleston CJ, Marks BD, Miglia KJ, Moore WS, Sheldon FH, Steadman DW, Witt CC, Yuri T (2008) A phylogenomic study of birds reveals their evolutionary history. Science 320:1763–1768
- Harvey PH, Pagel MD (1991) The comparative method in evolutionary biology. Oxford University Press, Oxford

- Honza M, Prochazka P, Stokke BG, Moksnes A, Røskaft E, Capek M, Mrlik V (2004) Are blackcaps current winners in the evolutionary struggle against the common cuckoo? J Ethol 22:175–180
- Houston CS, Saunders JR, Crawford RD (1997) Aerobic bacterial flora of addled raptor eggs in Saskatchewan. J Wildl Dis 33:328–331
- Iwaniuk AN, Nelson JE (2003) Developmental differences are correlated with relative brain size in birds: a comparative analysis. Can J Zool 81:1913–1928
- Järvinen A (1984) Relationship between the common cuckoo *Cuculus canorus* an its host, the redstart *Phoenicurus phoenicurus*. Ornis Fenn 61:84–88
- Jønsson KA, Fjeldså J (2006) A phylogenetic supertree of oscine passerine birds (Aves: Passeri). Zool Scripta 35:149–186
- Krieg NR, Holt JG (1984) Bergey's manual of systematic bacteriology. Williams & Wilkins, Baltimore, Maryland
- Lefebvre L, Bolhuis JJ (2003) Positive and negative correlates of feeding innovation in birds: evidence for limited modularity. In: Reader SM, Laland KN (eds) Animal innovation. Oxford University Press, Oxford, pp 39–61
- Lefebvre L, Whittle P, Lascaris E, Finkelstein A (1997) Feeding innovations and forebrain size in birds. Anim Behav 53:549–560
- Lefebvre L, Juretic N, Nicolakakis N, Timmermans S (2001) Is the link between forebrain size and feeding innovations caused by confounding variables? A study of Australian and North American birds. Anim Cogn 4:91–97
- Lefebvre L, Nicolakakis N, Boire D (2002) Tools and brains in birds. Behaviour 139:939–973
- Lefebvre L, Reader SM, Sol D (2004) Brains, innovations and evolution in birds and primates. Brain Behav Evol 63:233–246
- Lindholm AK, Thomas RJ (2000) Differences between populations of reed warblers in defences against brood parasitism. Behaviour 137:25–42
- Lotem A (1993) Learning to recognize nestling is maladaptive for cuckoo *Cuculus canorus* host. Nature 362:743–745
- Lotem A, Nakamura H, Zahavi A (1995) Constraints on egg discrimination and cuckoo host coevolution. Anim Behav 49:1185–1209
- Lovaszi P, Moskat C (2004) Break-down of arms race between the redbacked shrike (*Lanius collurio*) and common cuckoo (*Cuculus canorus*). Behaviour 141:245–262
- Lozano GA (1998) Parasitic stress and self-medication in wild animals. Adv Stud Behav 27:291–317
- Lyon BE (2003) Egg regognition and counting reduce costs of avian conspecific brood parasitism. Nature 422:495–499
- Mallon EB, Brockmann A, Schmid-Hempel P (2003) Immune response inhibits associative learning in insects. Proc R Soc Lond B 270:2471–2473
- Martin TE, Møller AP, Merino S, Clobert J (2001) Does clutch size evolve in response to parasites and immunocompetence? Proc Natl Acad Sci USA 98:2071–2076
- Martín-Vivaldi M, Soler M, Møller AP (2002) Unrealistically high costs of rejecting artificial model eggs in cuckoo *Cuculus canorus* hosts. J Avian Biol 33:295–301
- Martín-Vivaldi M, Ruiz-Rodríguez M, Mendez M, Soler JJ (2006) Relative importance of factors affecting nestling immune response differs between junior and senior nestlings within broods of hoopoes *Upupa epops*. J Avian Biol 37:467–476
- Martín-Vivaldi M, Peña A, Peralta-Sánchez JM, Sánchez L, Ananou S, Ruiz-Rodríguez M, Soler JJ (2010) Antimicrobial chemicals in hoopoe preen secretions are produced by symbiotic bacteria. Proc R Soc B 277:123–130
- Martín-Vivaldi M, Soler JJ, Møller AP, Pérez-Contreras T, Soler M (2012) Importance of nest site and habitat in egg recognition ability of potential hosts of the European cuckoo *Cuculus canorus*. Ibis In press:

- Matson KD, Ricklefs RE, Klasing KC (2005) A hemolysis-hemagglutination assay for characterizing constitutive innate humoral immunity in wild and domestic birds. Dev Comp Imm 29:275–286
- McArthur JV (2006) Microbial ecology: an evolutionary approach. Elsevier Academic Press, Amsterdam
- Mennerat A, Mirleau P, Blondel J, Perret P, Lambrechts M, Heeb P (2009) Aromatic plants in nests of the blue tit *Cyanistes caeruleus* protect chicks from bacteria. Oecologia 161:849–855
- Merino S, Potti J (1996) Weather dependent effects of nest ectoparasites on their bird hosts. Ecography 19:107–113
- Mlikovsky J (1989) Brain size in birds: 2. Falconiformes through Gaviiformes. Vest cs Spolec Zool 53:200–213
- Moksnes A, Røskaft E, Braa AT, Korsnes L, Lampe HM, Pedersen HC (1991) Behavioural responses of potential hosts towards artificial cuckoo eggs and dummies. Behaviour 116:64–89
- Moksnes A, Røskaft E, Solli MM (1994) Documenting puncture ejection of parasitic eggs by chaffinches *Fringilla coelebs* and blackaps *Sylvia atricapilla*. Fauna Norvergica, Ser C, Cinclus 17:115–118
- Møller AP (1987) Egg predation as a selective factor for nest design: an experiment. Oikos 50:91–94
- Møller AP (1997) Parasitism and the evolution of host life history. In: Clayton DH, Moore J (eds) Host-parasite evolution: general principles and avian models. Oxford Univesity Press, Oxford, pp 105–127
- Møller AP, Erritzøe J (1998) Host immune defence and migration in birds. Evol Ecol 12:945–953
- Møller AP, Erritzøe J (2002) Coevolution of host immune defence and parasite-induced mortality: relative spleen size and mortality in altricial birds. Oikos 99:95–100
- Møller AP, Martín-Vivaldi M, Merino S, Soler JJ (2006) Densitydependent and geographical variation in bird immune response. Oikos 115:463–474
- Moreno J, Briones V, Merino S, Ballesteros C, Sanz JJ, Tomás G (2003) Beneficial effects of cloacal bacteria on growth and fledging size in nestling pied flycatchers (*Ficedula hypoleuca*) in Spain. Auk 120:784–790
- Moskat C, Fuisz TI (1999) Reactions of red-backed shrikes Lanius collurio to artificial cuckoo Cuculus canorus eggs. J Avian Biol 30:175–181
- Moskat C, Karcza Z, Csörgö T (2003a) Egg rejection in European blackbirds (*Turdus merula*): the effect of mimicry. Ornis Fenn 80:86–91
- Moskat C, Szekely T, Kisbenedek T, Karcza Z, Bartol I (2003b) The importance of nest cleaning in egg rejection behaviour of great reed warblers Acrocephalus arundinaceus. J Avian Biol 34:16–19
- Narushin VG (2005) Production, modeling, and education: egg geometry calculation using the measurements of length and breadth. Poultry Sci 84:482–484
- Nicolakakis N, Lefebvre L (2000) Forebrain size and innovation rate in European birds: feeding, nesting and confounding variables. Behaviour 137:1415–1429
- Nicolakakis N, Sol D, Lefebvre L (2003) Behavioural flexibility predicts species richness in birds, but not extinction risk. Anim Behav 65:445–452
- Niven JE, Laughlin SB (2008) Energy limitation as a selective pressure on the evolution of sensory systems. J Exp Biol 211:1792–1804
- Overington SE, Morand-Ferron J, Boogert NJ, Lefebvre L (2009) Technical innovations drive the relationship between innovativeness and residual brain size in birds. Anim Behav 78:1001–1010
- Pagel M (1997) Inferring evolutionary processes from phylogenies. Zool Scripta 26:331–348
- Pagel M (1999) Inferring the historical patterns of biological evolution. Nature 401:877–884
- Palomino JJ (1997) Reproductive ecology and parental care in the rufous-tailed scrub-robin: responses to parasitism by cuckoo. PhD thesis University of Granada, Granada.

- Payne RB (1977) The ecology of brood parasitism in birds. Ann Rev Ecol Syst 8:1–28
- Peralta-Sánchez JM (2011) Las bacterias como agentes modeladores de las estrategias vitales en aves. PhD thesis University of Granada, Granada.
- Peralta-Sánchez JM, Møller AP, Martín-Platero AM, Soler JJ (2010) Number and colour composition of nest lining feathers predict eggshell bacterial community in barn swallow nests: an experimental study. Funct Ecol 24:426–433
- Playfair J, Bancroft G (2004) Infection and immunity. Oxford University Press, Oxford
- Pons JM, Hassanin A, Crochet PA (2005) Phylogenetic relationships within the Laridae (Charadriiformes: Aves) inferred from mitochondrial markers. Mol Phylogenet Evol 37:686–699
- Prochazka P, Honza M (2003) Do common whitethroats (Sylvia communis) discriminate against alien eggs? J Ornithol 144:354–363
- Prochazka P, Honza M (2004) Egg discrimination in the yellowhammer. Condor 106:405–410
- Riddell CE, Mallon EB (2006) Insect psychoneuroimmunology: immune response reduces learning in protein starved bumblebees (*Bombus terrestris*). Brain Behav Immun 20:135–138
- Rohwer S, Spaw CD, Røskaft E (1989) Cost to northern orioles of puncture-ejecting parasitic cowbird eggs from their nest. Auk 106:734–738
- Rothstein SI (1975a) An experimental and teleonomic investigation of avian brood parasitism. Condor 77:250–271
- Rothstein SI (1975b) Mechanisms of avian egg-recognition: do birds know their own eggs? Anim Behav 23:268–278
- Rothstein SI (1990) A model system for coevolution: avian brood parasitism. Ann Rev Ecol Syst 21:481–508
- Ruiz-Gonzalez MX, Brown MJF (2006) Honey bee and bumblebee trypanosomatids: specificity and potential for transmission. Ecol Entomol 31:616–622
- Ruiz-Rodriguez M, Lucas FS, Heeb P, Soler JJ (2009) Differences in intestinal microbiota between avian brood parasites and their hosts. Biol J Linn Soc 96:406–414
- Rutila J, Latja R, Koskela K (2002) The common cuckoo Cuculus canorus and its cavity nesting host, the redstart Phoenicurus phoenicurus: a peculiar cuckoo–host system? J Avian Biol 33:414–419
- Rutila J, Jokimäki J, Aviles JM, Kaisanlahti-Jokimaki ML (2006) Responses of parasitized and unparasitized common redstart (*Phoenicurus phoenicurus*) populations against artificial cuckoo parasitism. Auk 123:259–265
- Sæther SA, Grøtan V, Engen S, Noble DG, Freckleton RP (2011) Rarity, life history and scaling of the dynamics in time and space of British birds. J Anim Ecol 80:215–224
- Sengupta B, Stemmler M, Laughlin SB, Niven JE (2010) Action potential energy efficiency varies among neuron types in vertebrates and invertebrates. PLoS Computational Biology 6:e1000840
- Shizuka D, Lyon BE (2010) Coots use hatch order to learn to recognize and reject conspecific brood parasitic chicks. Nature 463:223– U108
- Singleton DR, Harper RG (1998) Bacteria in old house wren nests. J Field Ornithol 69:71–74
- Sol D, Lefebvre L (2000) Behavioural flexibility predicts invasion success in birds introduced to New Zealand. Oikos 90:599–605
- Sol D, Timmermans S, Lefebvre L (2002) Behavioural flexibility and invasion success in birds. Anim Behav 63:495–502
- Sol D, Duncan RP, Blackburn TM, Cassey P, Lefebvre L (2005) Big brains, enhanced cognition, and response of birds to novel environments. Proc Natl Acad Sci USA 102:5460–5465
- Soler JJ (1999) Do life-history variables of European cuckoo hosts explain their egg-rejection behavior? Behav Ecol 10:1–6
- Soler JJ, Avilés JM (2010) Sibling competition and conspicuousness of nestling gapes in altricial birds: a comparative study. PLoS One 5: e10509

- Soler M, Møller AP (1990) Duration of sympatry and coevolution between the great spotted cuckoo and its magpie host. Nature 343:748–750
- Soler JJ, Møller AP (1996) A comparative analysis of the evolution of variation in appearance of eggs of European passerines in relation to brood parasitism. Behav Ecol 7:89–94
- Soler JJ, Martínez JG, Soler M, Møller AP (1999a) Genetic and geographic variation in rejection behavior of cuckoo eggs by European magpie populations: an experimental test of rejectergene flow. Evolution 53:947–956
- Soler JJ, Møller AP, Soler M (1999b) A comparative study of host selection in the European cuckoo *Cuculus canorus*. Oecologia 118:265–276
- Soler M, Martín-Vivaldi M, Pérez-Contreras T (2002) Identification of the sex responsible for recognition and the method of ejection of parasitic eggs in some potential common cuckoo hosts. Ethology 108:1093–1101
- Soler JJ, Martín-Vivaldi M, Ruiz-Rodríguez M, Valdivia E, Martín-Platero AM, Martínez-Bueno M, Peralta-Sánchez JM, Méndez M (2008) Symbiotic association between hoopoes and antibiotic-producing bacteria that live in their uropygial gland. Funct Ecol 22:864–871
- Soler JJ, Martín-Vivaldi M, Peralta-Sánchez JM, Ruiz-Rodríguez M (2010) Antibiotic-producing bacteria as a possible defence of birds against pathogenic microorganisms. Open Ornithol J 3:93–100
- Soler JJ, Peralta-Sánchez JM, Martínez Bueno M, Martín-Vivaldi M, Martín-Gálvez D, Vela AI, Briones V, Pérez-Contreras T (2011)

Brood parasitism is associated with increased bacterial contamination of host eggs: bacterial loads of host and parasitic eggs. Biol J Linn Soc 103:836–848

- Stadelman WJ (1994) Contaminats of liquid eggs products. In: Board RG, Fuller R (eds) Microbiology of the avian egg. Chapman & Hall, London, pp 139–151
- Stokke BG, Moksnes A, Røskaft E, Rudolfsen G, Honza M (1999) Rejection of artificial cuckoo (*Cuculus canorus*) eggs in relation to variation in egg appearance among reed warblers (*Acrocephalus scirpaceus*). Proc R Soc Lond B 266:1483–1488
- Stokke BG, Hafstad I, Rudolfsen G, Moksnes A, Møller AP, Røskaft E, Soler M (2008) Predictors of resistance to brood parasitism within and among reed warbler populations. Behav Ecol 19:612– 620
- Timmermans S, Lefebvre L, Boire D, Basu P (2000) Relative size of the hyperstriatum ventrale is the best predictor of feeding innovation rate in birds. Brain Behav Evol 56:196–203
- Tucker GM, Heath MF (1994) Birds in Europe, their conservation status. BirdLife International, Cambridge
- Vas Z, Lefebvre L, Johnson KP, Reiczigel J, Rózsa L (2011) Clever birds are lousy: co-variation between avian innovation and the taxonomic richness of their amblyceran lice. Int J Parasitol 41:1295–1300
- von Haartman L (1981) Co-evolution of the cuckoo *Cuculus canorus* and a regular cuckoo host. Ornis Fenn 58:1–10